

MicroRNA-93 Targets p21 and Promotes Proliferation in Mycosis Fungoides T Cells

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Keywords

Cutaneous T-cell lymphoma · Mycosis fungoides · microRNA-93 · p21 · Tumor progression · SAHA/Vorinostat

Abstract

Background: Mycosis fungoides (MF), the most common form of cutaneous T-cell lymphoma (CTCL), is a lymphoproliferative disorder characterized by proliferation of malignant T cells in a chronic inflammatory environment in the skin. The nature of MF is still not fully understood, but aberrant microRNA (miR) expression and function seem to play an important role in the pathogenesis and disease progression and have been proposed as a putative disease marker. Recent studies have reported aberrant expression of miR-93 in situ in MF lesions and linked dysregulated miR-93 expression to advanced stages of MF. However, the pathophysiological role of miR-93 in MF is unknown. **Objective:** Here, we provide the first evidence that miR-93 targets the cell cycle regulator cyclin-dependent kinase inhibitor p21 and promotes growth of malignant T cells in MF. **Methods/Results:** Thus, inhibition of miR-93 in MF patient-derived malignant T-cell lines increases expression of p21 and inhibition of malignant proliferation. Notably, treatment with the histone

deacetylase inhibitor Vorinostat (SAHA) reduces miR-93 expression and enhances p21 expression in the malignant T cells. Importantly, transfection with an miR-93 mimic partly blocks SAHA-induced p21 expression. **Conclusions:** we provide evidence that enhanced expression of the putative oncogenic miR, miR-93, represses the cell cycle inhibitor p21 and promotes proliferation of malignant T cells. Moreover, we demonstrate that SAHA triggers p21 expression – at least partly – through an inhibition of miR-93.

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Introduction

Cutaneous T-cell lymphoma (CTCL) represents a heterogeneous group of extranodal non-Hodgkin lymphoproliferative disorders arising from abnormal T cells and is characterized by cutaneous infiltrates of malignant T cells [1, 2]. The most common variant mycosis fungoides (MF) arises from skin-resident memory T cells [3]. The etiology remains unknown, and genetic, epigenetic,

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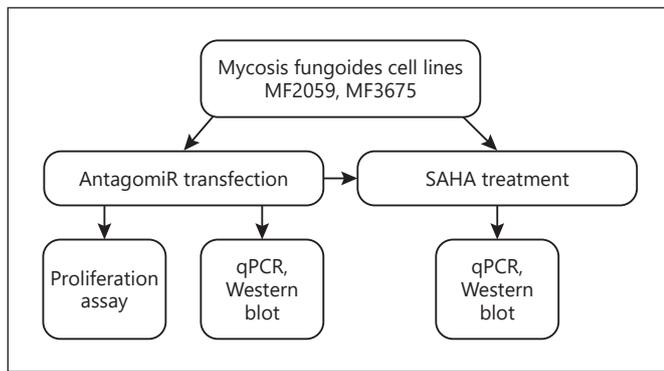


Fig. 1. Flowchart of Materials and Methods.

and environmental factors have all been implicated in CTCL and probably contribute to the high level of disease heterogeneity [4–6]. Notably, a recent study on CTCL in a Danish cohort of twins showed no evidence for heredity playing a role in CTCL [7]. In parallel, emerging evidence suggests that environmental factors may have both an etiological and a pathogenic role in CTCL [8–11]. Importantly, the pathogenesis of CTCL including Sézary syndrome (SS) is closely associated with chronic inflammation and aberrant activation of the NF- κ B pathway, NFAT pathway, and JAK/STAT pathway in lesional skin and blood [12–15]. Dysregulation of microRNA molecules (miRs) has also been implicated in the pathogenesis of CTCL [2, 16–19]. miRs are small noncoding RNAs consisting of 20–22 nucleotides, which interfere with gene regulation at the post-transcriptional level and regulate crucial cellular processes including development, proliferation, and survival [20]. miR profiles in CTCL have been extensively studied and demonstrate unique profiles with diagnostic and prognostic potential, including the ability to discriminate between CTCL and benign skin diseases as well as different stages of CTCL [17, 19, 21, 22]. miR-155 is one of the best-characterized miRs in CTCL and is classified as an oncogenic miR (onco-miR). The high expression of miR-155 mediates inhibition of the tumor suppressor SATB1 and induction of proliferation in malignant T cells [16, 23]. In contrast, miR-22, miR-203, and miR-205 are significantly down-regulated and fail to execute their putative tumor suppressive functions in CTCL [22, 24]. Interestingly, several studies investigating miR profiles have demonstrated that miR-93 is highly overexpressed in MF [17, 19, 25] but not in SS [26]. miR-93 is clustered with miR-25 and miR-106b and located in intron 13 hosted by the gene encoding MCM7, which expression has been reported to

be elevated in MF [27, 28]. Studies have additionally shown that miR-93 is upregulated in advanced stages of MF, thus suggesting a role for this miR in disease progression [19]. Furthermore, miR-93 has previously been described in a range of cancers as an onco-miR preventing apoptosis and promoting tumor survival [29]. Despite its apparent dysregulation in CTCL, the role of miR-93 remains to be elucidated. Deregulation of numerous tumor suppressors, including the important cell cycle inhibitor p21, has been described in CTCL, and tumor suppressors may serve as potential targets for miR regulation [30]. In this study, we sought to gain further insights into the miR-93-mediated effects on tumor progression.

Materials and Methods

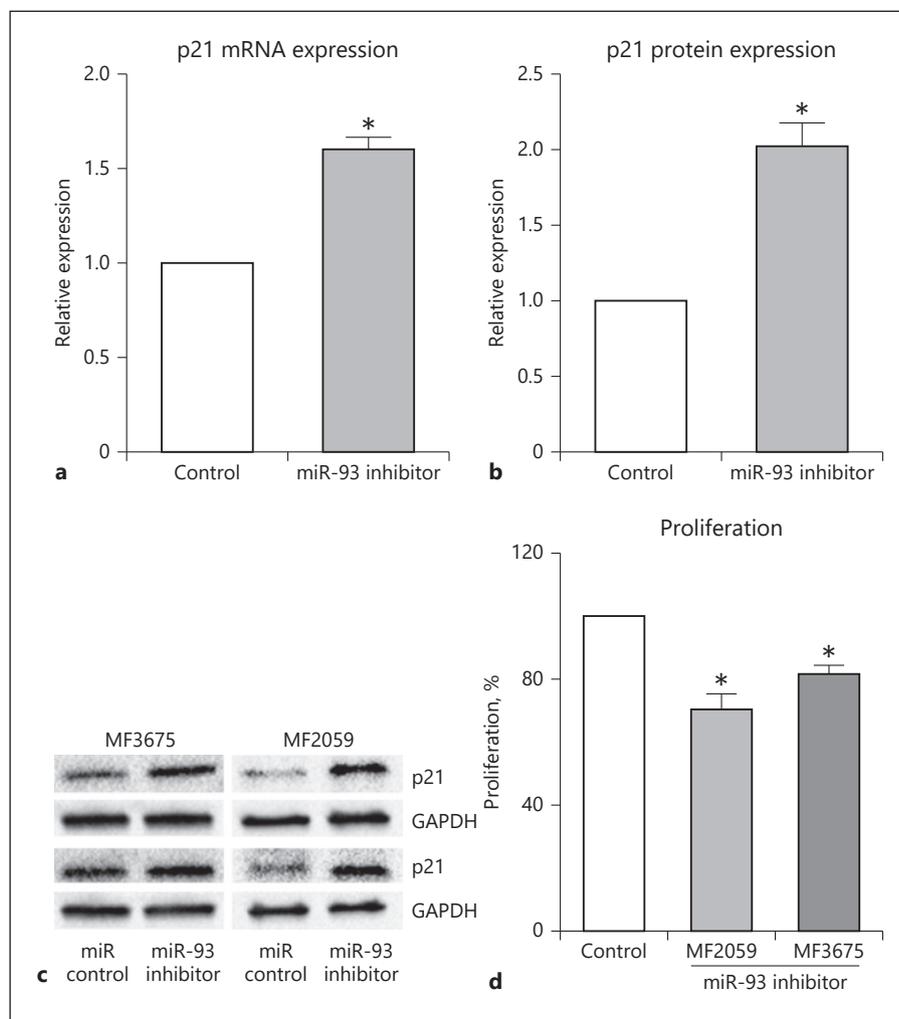
For further details, see online supplementary material (see www.karger.com/doi/10.1159/000505743 for all online suppl. material) [31, 32] (Fig. 1).

Results

In order to determine the role of miR-93, which is hypothesized to be involved in the progression of MF [19], the tumor cell lines MF2059 and MF3675 were transfected with an miR-93-5p inhibitor to reduce the activity of the miR. Following 24 h of incubation with the inhibitor, a 50% increase in p21 mRNA in MF2059 cells and a profound increase in the p21 protein expression in MF2059 and MF3675 cell lines were observed (Fig. 2a–c). However, the SS cell line SeAx did not show any inhibition of p21 protein expression following miR-93 inhibition (online suppl. Fig. S1). To determine the effect on proliferation, thymidine incorporation was measured following transfection experiments with miR-93 inhibitors. Inhibition of miR-93 resulted in a 20–30% reduced proliferation of the malignant T-cell lines (Fig. 2d). This reduced proliferation is comparable to the effects observed following inhibition of miR-155, which is a well-established miR known to play an important role in MF [16] (online suppl. Fig. S2).

The histone deacetylase (HDAC) inhibitor suberoylanilide hydroxamic acid (SAHA/Vorinostat), which modulates the epigenetic landscape of malignant T cells, is an approved treatment option for CTCL [33, 34]. In order to test the hypothesis that HDAC inhibitors exert their functions through regulation of miRs, SAHA was

Fig. 2. miR-93 downregulates the expression of p21 and reduces proliferation in malignant T cells. The tumor cell lines MF2059 and MF3675 were transfected with miR-93 inhibitors and cultured for 24 h prior to analysis. **a** mRNA expression of p21 was measured in MF2059 cells by qPCR ($n = 3$) using GAPDH as a reference gene. **b** Western blotting was used to determine the protein expression of p21 in MF2059 and the intensity of the bands were quantified ($n = 4$). **c** Western blotting analysis was evaluated for MF2059 and MF3675 cells ($n = 2$). GAPDH was applied as a loading control. **d** The MF2059 and MF3675 cells were transfected with miR-93 inhibitors and cultured for 24 h in the presence of ^3H -thymidine. Incorporation of ^3H -thymidine was analyzed to determine the proliferation of malignant T cells ($n = 4$). * $p < 0.05$, indicates statistical significance.



added in concentrations ranging from 0.6 to 5 μM to the culture of the malignant T-cell line MF2059. SAHA reduced the expression of primary miR-93 by up to 72% in a concentration-dependent manner (Fig. 3a). Furthermore, the addition of SAHA induced the expression of the tumor suppressor miR-22 primary transcript and the expression of p21 mRNA, consistent with previous reports (Fig. 3b, c) [35, 36]. Protein expression analyzed by Western blotting confirmed that SAHA enhanced p21 protein expression and that the reduction of miR-93 activity and SAHA additively induced the expression of p21 protein (Fig. 3d). As illustrated in Figure 3e, transfection experiments applying miR-93 mimics reduced the upregulation of p21 observed as a response to SAHA treatment (Fig. 3e), indicating that SAHA-induced p21 expression is mediated, at least partly, through an inhibition of miR-93.

Discussion/Conclusion

miRs play a key role in the regulation of T-cell development, differentiation, and proliferation [20]. Abnormal miR expression profiles have been strongly implicated in a broad range of cancers, and patients suffering from CTCL show distinct miR expression profiles, which may act to promote tumor progression [19, 21]. miR-93 has been reported to be significantly upregulated in advanced stages of MF, which suggests a role in the progression of the disease [17, 19]. However, miR-93 did not serve as a prognostic marker in a large cohort of early-stage MF patients [21]. Here we report that miR-93 interferes with p21 expression in MF tumor T-cell lines but not in an SS cell line. p21, a known cell cycle inhibitor and tumor suppressor, is one of many dysregulated tumor suppressors reported in CTCL [30]. We furthermore demonstrate

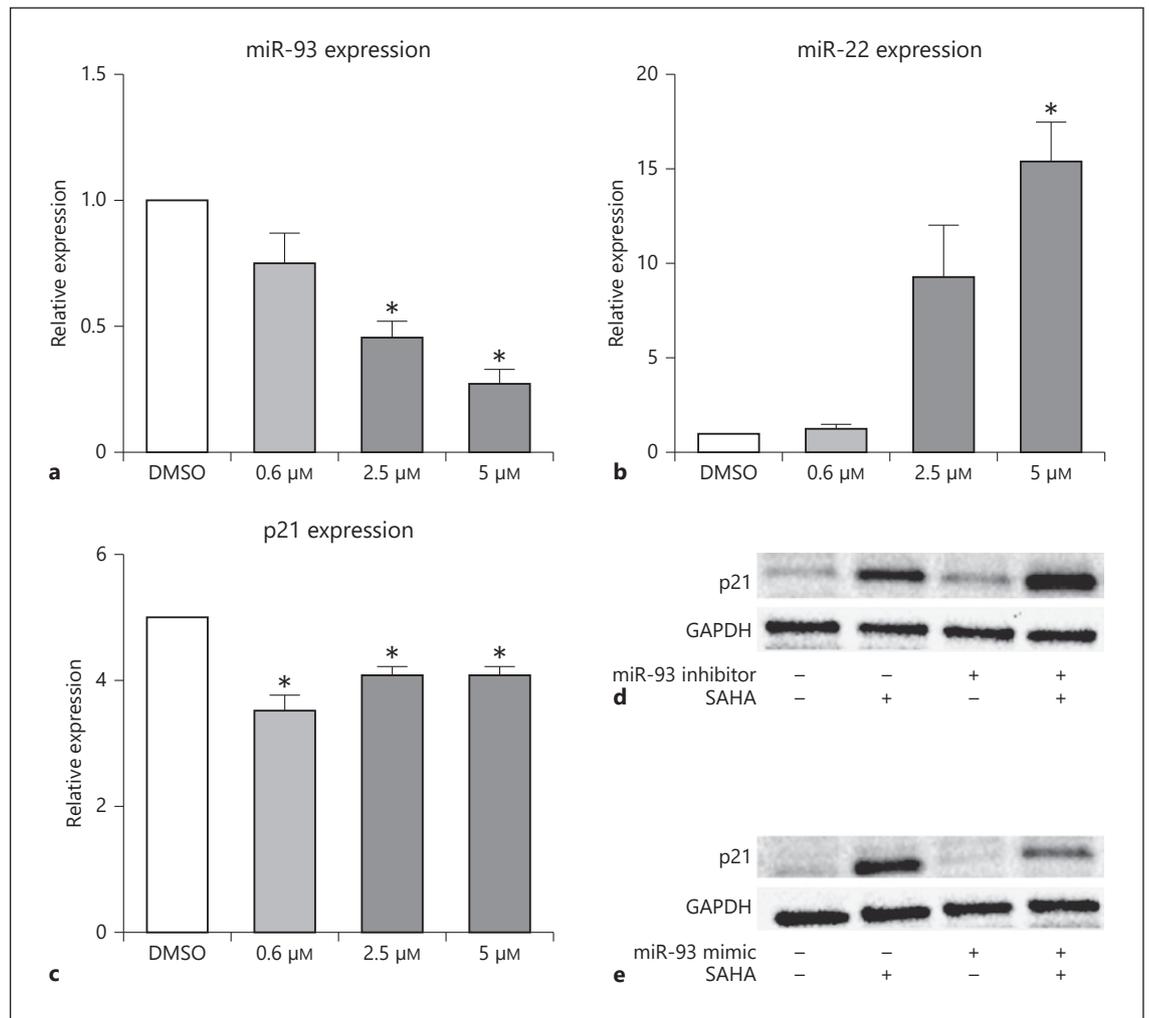


Fig. 3. SAHA treatment inhibits miR-93 and induces miR-22 and p21 in malignant T cells. The tumor cell line MF2059 was cultured for 24 h with concentrations of SAHA ranging from 0.6 to 5 μM prior to analysis. Primary miR levels were measured in MF2059 cells by qPCR for miR-93 (a) ($n = 3$), miR-22 (b) ($n = 3$), and mRNA expression for p21 (c) ($n = 3$) using GAPDH as a reference gene. Western blotting was used to evaluate the protein expression of p21 in MF2059 cells transfected with either miR-93 inhibitors and controls (d) or miR-93 mimics and controls (e). Cells were transfected and incubated for 24 h prior to being cultured for 24 h in the presence of SAHA or DMSO. GAPDH was applied as a loading control. Statistical significance was determined using one-way ANOVA, with multiple comparison tests. * $p < 0.05$, indicates statistical significance.

that miR-93 inhibition decreases the proliferation of malignant T cells, suggesting that the aberrant expression of miR-93 seen in MF may accelerate proliferation of malignant T cells. The effects on proliferation were comparable to those observed for miR-155 inhibition, a well-characterized onco-miR in CTCL, which currently is being investigated as a therapeutic target in clinical trials [37]. miRs regulate many genes post-transcriptionally, thus numerous miRs may act through multiple mechanisms to promote proliferation of malignant T cells [20]. A recent

study describes that miR-16 indirectly targets p21 in CTCL, thus contributing to the diminished expression of p21 [35]. This indicates that repression of p21 may be a key event in disease progression in MF and that multiple miRs, directly or indirectly, orchestrate an inhibition of p21 expression at the protein level.

Deciphering the molecular mechanisms leading to cancer progression is crucial in the search for new therapeutic targets as well as in the understanding of the underlying mechanisms of already existing therapeutics.

The mode of action of SAHA remains to be completely elucidated [34]. Studies have shown that SAHA upregulates p21, inhibits cell growth of malignant T cells, and induces apoptosis [38]. In the present study, we provide evidence of a possible mechanism by which SAHA exerts part of its effects, as SAHA reduces miR-93 expression which in turn influences the regulation of p21. Our findings are in concordance with previous studies which indicate that SAHA regulates numerous miRs [24, 35], downregulates tumor-promoting miRs, and induces important tumor suppressor miRs, such as miR-22. In conclusion, we provide novel insights into the role of miR-93 as a potential tumor-promoting factor in MF. Further characterization of potential mechanisms of action in relation to tumor promotion could advance our understanding of miR-93 and contribute to the establishment of miR-93 as an important factor in tumor progression. Thus, miR-93 remains a target of great interest, and further studies are required to determine the potential of miR-93 as a treatment target in MF.

Key Message

The highly expressed miR-93 decreases p21 expression and promotes proliferation in mycosis fungoides T cells.

References

- Kim EJ, Hess S, Richardson SK, Newton S, Showe LC, Benoit BM, et al. Immunopathogenesis and therapy of cutaneous T cell lymphoma. *J Clin Invest*. 2005 Apr;115(4):798–812.
- Wilcox RA. Cutaneous T-cell lymphoma: 2017 update on diagnosis, risk-stratification, and management. *Am J Hematol*. 2017 Oct;92(10):1085–102.
- Campbell JJ, Clark RA, Watanabe R, Kupper TS. Sézary syndrome and mycosis fungoides arise from distinct T-cell subsets: a biologic rationale for their distinct clinical behaviors. *Blood*. 2010 Aug;116(5):767–71.
- Litvinov IV, Shtreiss A, Kobayashi K, Glassman S, Tsang M, Woetmann A, et al. Investigating potential exogenous tumor initiating and promoting factors for Cutaneous T-Cell Lymphomas (CTCL), a rare skin malignancy. *OncoImmunology*. 2016 Jun;5(7):e1175799.
- Buus TB, Willerslev-Olsen A, Fredholm S, Blümel E, Nastasi C, Gluud M, et al. Single-cell heterogeneity in Sézary syndrome. *Blood Adv*. 2018 Aug;2(16):2115–26.
- Bastidas Torres AN, Najidh S, Tensen CP, Vermeer MH. Molecular advances in cutaneous T-cell lymphoma. *Semin Cutan Med Surg*. 2018 Mar;37(1):81–6.
- Odum N, Lindahl LM, Wod M, Krejsgaard T, Skyttte A, Woetmann A, et al. Investigating heredity in cutaneous T-cell lymphoma in a unique cohort of Danish twins. *Blood Cancer J*. 2017 Jan;7(1):e517.
- Willerslev-Olsen A, Krejsgaard T, Lindahl LM, Litvinov IV, Fredholm S, Petersen DL, et al. Staphylococcus aureus enterotoxin A (SEA) stimulates STAT3 activation and IL-17 expression in cutaneous T-cell lymphoma. *Blood*. 2016 Jan;127(10):1287–96.
- Ghazawi FM, Netchiporouk E, Rahme E, Tsang M, Moreau L, Glassman S, et al. Comprehensive analysis of cutaneous T-cell lymphoma (CTCL) incidence and mortality in Canada reveals changing trends and geographic clustering for this malignancy. *Cancer*. 2017 Sep;123(18):3550–67.
- Lindahl LM, Willerslev-Olsen A, Gjerdrum LM, Nielsen PR, Blümel E, Rittig AH, et al. Antibiotics inhibit tumor and disease activity in cutaneous T-cell lymphoma. *Blood*. 2019 Sep;134(13):1072–83.
- Blümel E, Willerslev-Olsen A, Gluud M, Lindahl LM, Fredholm S, Nastasi C, et al. Staphylococcal alpha-toxin tilts the balance between malignant and non-malignant CD4+ T cells in cutaneous T-cell lymphoma. *Oncoimmunology*. 2019 Jul;8(11):e1641387.
- Sors A, Jean-Louis F, Pellet C, Laroche L, Dubertret L, Courtois G, et al. Down-regulating constitutive activation of the NF-kappaB canonical pathway overcomes the resistance of cutaneous T-cell lymphoma to apoptosis. *Blood*. 2006 Mar;107(6):2354–63.
- Nielsen M, Nissen MH, Gerwien J, Zocca MB, Rasmussen HM, Nakajima K, et al. Spontaneous interleukin-5 production in cutaneous T-cell lymphoma lines is mediated by constitutively activated Stat3. *Blood*. 2002 Feb;99(3):973–7.
- Kiel MJ, Sahasrabudhe AA, Rolland DC, Velusamy T, Chung F, Schaller M, et al. Genomic analyses reveal recurrent mutations in epigenetic modifiers and the JAK-STAT pathway in Sézary syndrome. *Nat Commun*. 2015 Sep;6(1):8470.
- Krejsgaard T, Lindahl LM, Mongan NP, Wasik MA, Litvinov IV, Iversen L, et al. Malignant inflammation in cutaneous T-cell lymphoma—a hostile takeover. *Semin Immunopathol*. 2017 Apr;39(3):269–82.
- Kopp KL, Ralfkiaer U, Gjerdrum LM, Helvad R, Pedersen IH, Litman T, et al. STAT5-mediated expression of oncogenic miR-155 in cutaneous T-cell lymphoma. *Cell Cycle*. 2013 Jun;12(12):1939–47.

Statement of Ethics

In accordance with the Declaration of Helsinki, samples were obtained with written consent from the patients after approval by the Committee on Health Research.

Disclosure Statement

The authors have no conflicts of interest to declare.

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Author Contributions

M.G. performed the experiments. M.G., S.F., E.B., A.W.-O., T.B.B., C.N., and T.K. analyzed the data and made the figures. M.G., A.W., N.Ø., L.M.L., C.M.B., C.G., T.L., and L.I. designed the research. M.G. wrote the original draft of the manuscript. All authors reviewed and edited the manuscript. N.Ø. supervised the project and acquired the funding.

- 17 van Kester MS, Ballabio E, Benner MF, Chen XH, Saunders NJ, van der Fits L, et al. miRNA expression profiling of mycosis fungoides. *Mol Oncol*. 2011 Jun;5(3):273–80.
- 18 Lindahl LM, Fredholm S, Joseph C, Nielsen BS, Jønson L, Willerslev-Olsen A, et al. STAT5 induces miR-21 expression in cutaneous T cell lymphoma. *Oncotarget*. 2016 Jul;7(29):45730–44.
- 19 Ralfkiaer U, Lindahl LM, Litman T, Gjerdrum LM, Ahler CB, Gniadecki R, et al. MicroRNA expression in early mycosis fungoides is distinctly different from atopic dermatitis and advanced cutaneous T-cell lymphoma. *Anticancer Res*. 2014 Dec;34(12):7207–17.
- 20 Rupaimoole R, Slack FJ. MicroRNA therapeutics: towards a new era for the management of cancer and other diseases. *Nat Rev Drug Discov*. 2017 Mar;16(3):203–22.
- 21 Lindahl LM, Besenbacher S, Rittig AH, Celis P, Willerslev-Olsen A, Gjerdrum LM, et al. Prognostic miRNA classifier in early-stage mycosis fungoides: development and validation in a Danish nationwide study. *Blood*. 2018 Feb;131(7):759–70.
- 22 Ralfkiaer U, Hagedorn PH, Bangsgaard N, Løvendorf MB, Ahler CB, Svensson L, et al. Diagnostic microRNA profiling in cutaneous T-cell lymphoma (CTCL). *Blood*. 2011 Nov;118(22):5891–900.
- 23 Fredholm S, Willerslev-Olsen A, Met Ö, Kubat L, Gluud M, Mathiasen SL, et al. SATB1 in malignant T cells. *J Invest Dermatol*. 2018 Aug;138(8):1805–15.
- 24 Sibbesen NA, Kopp KL, Litvinov IV, Jønson L, Willerslev-Olsen A, Fredholm S, et al. Jak3, STAT3, and STAT5 inhibit expression of miR-22, a novel tumor suppressor microRNA, in cutaneous T-Cell lymphoma. *Oncotarget*. 2015 Aug;6(24):20555–69.
- 25 Marosvári D, Téglási V, Csala I, Marschalkó M, Bődör C, Timár B, et al. Altered microRNA expression in folliculotropic and transformed mycosis fungoides. *Pathol Oncol Res*. 2015 Jul;21(3):821–5.
- 26 Ballabio E, Mitchell T, van Kester MS, Taylor S, Dunlop HM, Chi J, et al. MicroRNA expression in Sezary syndrome: identification, function, and diagnostic potential. *Blood*. 2010 Aug;116(7):1105–13.
- 27 Jankowska-Konsur A, Kobierzycki C, Reich A, Grzegorzolka J, Maj J, Dziegiel P. Expression of MCM-3 and MCM-7 in Primary Cutaneous T-cell Lymphomas. *Anticancer Res*. 2015 Nov;35(11):6017–26.
- 28 Gambichler T, Bischoff S, Bechara FG, Altmeyer P, Kreuter A. Expression of proliferation markers and cell cycle regulators in T cell lymphoproliferative skin disorders. *J Dermatol Sci*. 2008 Feb;49(2):125–32.
- 29 Li N, Miao Y, Shan Y, Liu B, Li Y, Zhao L, et al. MiR-106b and miR-93 regulate cell progression by suppression of PTEN via PI3K/Akt pathway in breast cancer. *Cell Death Dis*. 2017 May;8(5):e2796.
- 30 Bagherani N, Smoller BR. An overview of cutaneous T cell lymphomas. *F1000Res*. 2016 Jul;5:F1000 Faculty Rev-1882.
- 31 Woetmann A, Lovato P, Eriksen KW, Krejsgaard T, Labuda T, Zhang Q, et al. Nonmalignant T cells stimulate growth of T-cell lymphoma cells in the presence of bacterial toxins. *Blood*. 2007 Apr;109(8):3325–32.
- 32 Fredholm S, Gjerdrum LMR, Willerslev-Olsen A, Petersen DL, Nielsen IØ, Kauczok C-S, et al. STAT3 Activation and infiltration of eosinophil granulocytes in mycosis fungoides. *Anticancer Res*. 2014 Jan;34(10):5277–86.
- 33 Zhang C, Richon V, Ni X, Talpur R, Duvic M. Selective induction of apoptosis by histone deacetylase inhibitor SAHA in cutaneous T-cell lymphoma cells: relevance to mechanism of therapeutic action. *J Invest Dermatol*. 2005 Nov;125(5):1045–52.
- 34 Lopez AT, Bates S, Geskin L. Current Status of HDAC Inhibitors in Cutaneous T-cell Lymphoma. *Am J Clin Dermatol*. 2018 Dec;19(6):805–19.
- 35 Kitadate A, Ikeda S, Teshima K, Ito M, Toyota I, Hasunuma N, et al. MicroRNA-16 mediates the regulation of a senescence-apoptosis switch in cutaneous T-cell and other non-Hodgkin lymphomas. *Oncogene*. 2016 Jul;35(28):3692–704.
- 36 Gui CY, Ngo L, Xu WS, Richon VM, Marks PA. Histone deacetylase (HDAC) inhibitor activation of p21WAF1 involves changes in promoter-associated proteins, including HDAC1. *Proc Natl Acad Sci USA*. 2004 Feb;101(5):1241–6.
- 37 Querfeld C, Foss FM, Pinter-Brown LC, Porcu P, William BM, Pacheco T, et al. Phase 1 Study of the Safety and Efficacy of MRG-106, a Synthetic Inhibitor of microRNA-155, in CTCL Patients. *Blood*. 2017 Dec;130 Suppl 1:820.
- 38 Al-Yacoub N, Fecker LF, Möbs M, Plötz M, Braun FK, Sterry W, et al. Apoptosis induction by SAHA in cutaneous T-cell lymphoma cells is related to downregulation of c-FLIP and enhanced TRAIL signaling. *J Invest Dermatol*. 2012 Sep;132(9):2263–74.